ISSN 2349-7041

Vol 8, Issue 3, May-Jun 2021

Research Article

ETHANOL INTOXICATED HEPATIC OXIDATIVE STRESS MITIGATED BY POLY-HERBAL FORMULATION – TRASINA® IN MURINE MODEL

Hepatic Oxidative Stress Prevented by Trasina®

SOUMENDRA DARBAR^{1*}, SRIMOYEE SAHA² AND ATISKUMAR CHATTOPADHYAY³

¹Research and Development Division, Dey's Medical Stores (Mfg.) Ltd., 62, Bondel Road, Kolkata-700019, West Bengal, India

²Department of Physics, Jadavpur University, 188, Raja S C Mallick Road, Kolkata-700032, West Bengal, India ³Faculty Council of Science, Jadavpur University, 188, Raja S C Mallick Road, Kolkata-700032, West Bengal, India

*Email: dr.soumendradarbar@deysmedical.com

ABSTRACT

Background: The main focus and aim of this scientific novel work was to find out the probable ameliorative effect of poly-herbal formulation (Trasina®) on serum and liver antioxidant enzymes activities in ethanol intoxicated organ dysfunctions in mice model.

Methods: Forty Swiss albino adult male mice were taken from our animal house and randomly break into 4 groups; Group-1 served as control, Group-2 orally treated with ethanol (50% v/v), Group-3 pre-treated with herbal medicine (Trasina®) along with ethanol (50% v/v), and Group-4 only treated with poly-herbal formulation (Trasina®) without ethanol daily. Completion of six weeks treatment the animals were euthanized and livers were removed immediately and used fresh or kept frozen until analysis. Blood was taken from the animals before sacrifice for measurement the antioxidant parameters first and second order enzymes i.e. catalase (CAT), super oxide dismutase (SOD), glutathione – S transferees (GST) and reduce glutathione (GSH) from sera.

Results: Activities of all antioxidant enzymes i.e. SOD, CAT, GSH and GST in serum and liver were significantly decreased in the ethanol intoxicated mice than in the controls. Treatment with herbal medicine (Trasina[®]) upon ethanol intoxication significant elevated the all antioxidant activities serum and liver.

Conclusions: Results obtained from the present study clearly predict that treatment with poly-herbal formulation (Trasina®) might be a potent antioxidant that exerts beneficial effects on both catalase (CAT), super oxide dismutase (SOD), glutathione –S transferees (GST) and glutathione peroxidase (GPx) activities in ethanol intoxicated mice and inhibit organ damage.

Keywords: Antioxidant enzymes activity, Liver, Ethanol, Trasina®, Oxidative stress, Swiss albino mice

INTRODUCTION

Mammalian normal cellular activity and physiology was severely affected if body attacks by Reactive oxygen species (ROS) [1-4]. During ethanol induced organ toxicity antioxidants play a crucial role for preventing the cellular damage caused by various redox molecules [5,6]. Commonly most of the xenobiotics were largely metabolised in the liver which reduced the toxic effects of the molecules. Most of the time by-products of this type of metabolism makes sever harmful effects and produce cellular imbalance [7]. This could lead liver damage and emergence hepatic disorders. In very frequent oxygen containing by-product molecules damage liver cell through oxidation. They produce oxidative stress and generates enormous amount of free radicals which creates membrane damage by free radicals, damage the membranous lipid and protein as well as modified deoxy ribonucleic acid (DNA) [8]. The generation of reactive oxygen species (ROS) and the defensive ability to counteract this deleterious effects was breaks by the ethanol which decline antioxidants functions [9]. Enormous formation and insufficient release of free radicals lead to severe and irreversible cell damage [10]. Scientific research work reviled that when cell attacked by free radicals through oxidative damage notably decrease in various antioxidant enzymes activities such as catalase (CAT), superoxide dismutase, glutathione S-transferase (GST), and reduce glutathione (GSH) which acts as a free radical scavengers in conditions associated with oxidative stress [11,12].

Indian systems of medicine have very safe and effective for curing various diseases with no toxic effects. Now a days throughout the world people relies on herbal drugs because of its less side effects in comparison to modern synthetic medicines. Most of the chronic diseases when treated with natural medicines considerably very useful for the effective medication. Prolonged used of the herbal medicine not showed adverse side effects and reaction because its contains lots of constituents those are highly medicinal property [13,17].

Trasina® a marketed poly-herbal capsule composed of five Indian medicinal plants consistently used from the ancient time for the benefit of mankind. Withania somnifera, Ocimum sanctum, Tinospora cordifolia, Picrorrhiza kurroa, Eclipta alba, and Shilajit, are the main ingredients present in Trasina® [18]. In 1997 Bhattacharya et al. reported that the said formulation has facilitating the memory action in rodent and nonrodent. Twenty one days sub chronic administration of Trasina on two different animal models revelled that this medicine had simulate some biochemical features known to be associated with Alzheimer's disease (AD) [19]. Our recent study confirmed that Trasina® has no toxic effects of animals and safe for therapeutic medication. Another very recent study established that Trasina® possessed significant antistress activity and maintain normal homeostasis [20,21]. This study stated that administration of Trasina® significantly increases anoxia tolerance time, significantly decreases immobility time and number of writhes in animals. Immobilisation stress induced changes in biochemical parameters and organs weight were completely revert by the application of Trasina® in experimental animals [18].

Thus, the present study was designed to assess the serum and liver antioxidant activity of poly-herbal capsule – Trasina® against ethanol induced oxidative stress in mice.

MATERIALS AND METHODS

Drugs and Chemicals

The poly-herbal capsule Trasina®, was taken from Dey's Medical (Kolkata, India). Ethanol, TRIS buffer and phosphate buffer were obtained from Merck, India. All antioxidant enzyme study kits were purchase from Emark Germany. All others necessary chemicals and reagents were procured from local renounced sources and were of analytical grade.

Animals

Forty young swiss albino male mice weighing 26–28 g were randomly chosen and used for the experiment. The animals have been kept with our well ventilated Animal House with all essential environment based on the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA) guidelines. The animals were acclimatized for 7 consecutive day's 12 h light and dark cycle. The animals were preserved in stainless steel (SS) cages with maintaining hygienic condition. The room was air-conditioned with temperature and humidity maintained. Mice were allowed standard chow diet (Amrut feeds, Pranav Agro, New Delhi, India) throughout the experimental period and water ad libitum. The entire experimental procedures were scrutinised and approved by the Institutional Animal Ethics Committee (IAEC) (Approval No. 16/IAEC/Dey's/s/2016).

Experimental design

Healthy adult male mice were divided into four experimental groups. Cage of the each group contain ten mice The details of the group division and treatment protocols are: Group 1 received vehicle and served as a control, Group 2 received 0.5 ml of Ethanol (50% v/v), Group 3 received Ethanol (50% v/v) along with Poly-Herbal formulation (Trasina®) (200 mg/kg) and Group 4 received only Poly-Herbal formulation (Trasina®) (200 mg/kg) for 6 weeks.

Sample collection

After completion of the experimental period, blood sample for analysis was collected from the retro orbital plexus. Collected blood samples stay for 1 h in normal room temperature ($27 \pm 2^{\circ}$ C) and then centrifuge at 6500 rpm for 15min to obtain clear serum. Serum was stored in aliquots at -60°C till used for estimation of various antioxidant enzymes.

After collection of blood sample, the abdomen and the thorax of the mice were opened and removed liver with proper care. The liver was washed four times in ice cold phosphate buffer saline and blotted individually on what man filter paper. The samples were then taken for homogenization for estimation of tissue antioxidant enzymes like SOD, CAT, GSH and GST.

Preparation of tissue homogenates

Small portion of liver was homogenization with a potter- Elvenhjem tissue homogenizer. Tissue was taken in phosphate buffer saline (PBS) 50 mM pH (7.4) as a homogenised medium. After the homogenization

aliquot was stores for the for estimation of total protein content, SOD, CAT, GST, GSH enzymes activities.

Determination of protein content

Serum and liver total protein levels was measured by the method of Kashyap *et al.* (Lowry assay) [22].

Determination of Lipid peroxidation

Lipid peroxidation (LPO) was measurement by using lipid peroxidation (MDA) assay kit (Sigma-Aldrich Ltd., UK) in accordance to the manufacturer's instructions. Broadly lipid peroxidation is detected by the reaction of malondialdehyde (MDA) with thiobarbituric acid (TBA) to form a colorimetric product, proportional to the MDA present. After addition of MDA-TBA adduct each sample was incubated at 90°C for 60 min, prior to cool to room temperature in an ice bath for 10 min. To conduct the reaction 200 mL mixture was transferred into a 96-well plate for analysis. The absorbance was measured at 532 nm [23].

Assay of antioxidant enzyme activities

a. Determination of Catalase (CAT) activity

Catalase (CAT) activity was measured by the method of Beutler *et al.* 1984 [24]. In brief, to a phosphate buffer (pH 7.0) and sample were added within a quartz cuvette. The reaction was started by addition of H_2O_2 . The decomposition of H_2O_2 was monitored at 240 nm.

b. Determination of Super oxide dismutase (SOD) activity

Activity of super oxide dismutase (SOD) in serum and hepatic tissue homogenate was assessed according to method of Misra *et al.* 1972 [25] with slight modification. In details 1 mL of Tris-HCl buffer, containing diethylene triaminopentaacetic acid and pyrogallol were mixed with 20 μ L of liver samples. The absorbance was measured at 440 nm.

c. Determination of Glutathione –S transferees (GST) activity

GST activity of Serum and hepatic tissues was investigated by the method of Beutler *et al.*1963 [26] with slight modification. The enzymatic reaction of 1chloro-2-4-di-nitrobenzene is neutralized by the enzyme in the presence of glutathione as a co substrate. The absorbance change was measured at 340 nm.

d. Determination of Reduce Glutathione (GSH) activity

Reduced glutathione (GSH) activity determination was based on the method of Jollow *et al.* 1974 [27]. In the

main reaction 1,2-dithio-bis nitro benzoic acid (DTNB) used as substrate. After adding the substrate the yellow color developed which was immediately read at 412 nm. The activity was expressed as μ mol GSH/g tissue.

Statistical analysis

SPSS (version 20.0) software were used for statistical. Tukey's test were used to determine significant differences between groups with one-way analysis of variance (ANOVA; P < 0.05). The values were stated as mean ± SD.

RESULTS

Protective effect of Poly-herbal medicine (Trasina®) on serum and hepatic protein content in ethanol toxicity

Poly-herbal medicine - Trasina[®] composed of five Indian medicinal plants (Table 1 and Figure 1) have been effective for stress and depression. Serum and hepatic protein contain are depicted in Table 2. Serum total protein content in the ethanol intoxicated mice was significantly lower than that of the controls $(3.62\pm0.16 \text{ vs. } 7.52\pm0.24 \text{ nmol/g})$. Pre-treatment with Poly-herbal medicine (Trasina[®]) significantly elevated Serum total protein content compared with ethanol intoxicated mice $(7.02\pm0.18 \text{ vs. } 3.62\pm0.16 \text{ nmol/g})$.

Hepatic total protein content in the ethanol intoxicated mice was significantly lower than that of controls $(2.05\pm0.09 \text{ vs } 5.92\pm0.11 \text{ nmol/g})$. Pre-treatment with Poly-herbal medicine (Trasina®) significantly decreased (Table 2) liver total protein content compared with ethanol intoxicated mice $(5.09\pm0.18 \text{ vs.} 2.05\pm0.09 \text{ nmol/g})$.

Protective effect of Poly-herbal medicine (Trasina®) on serum and liver MDA content in ethanol toxicity

Serum and tissue (liver) lipid peroxidation (MDA levels) are depicted in Table 3. Serum MDA content in the ethanol intoxicated mice was significantly higher than that of the controls (102.58±1.87 vs. 45.92±1.54 nmol/g). Pre-treatment with Poly-herbal medicine (Trasina®) significantly reduced Serum MDA content compared with ethanol intoxicated mice (49.63±0.91vs. 102.58±1.87 nmol/g).

Hepatic MDA content in the ethanol intoxicated mice was significantly high than that of controls (77.05±0.79 vs 35.28±0.92 nmol/g). Pre-treatment with Poly-herbal medicine (Trasina[®]) significantly decreased (Table 3) liver MDA content compared with ethanol intoxicated mice (39.01±0.88vs. 77.05±0.79 nmol/g).

Protective effect of Poly-herbal medicine (Trasina®) on serum and liver SOD activity in ethanol toxicity

Serum, and liver super oxide dismutase (SOD) activities are depicted in Figure 2. Serum activity of SOD in the ethanol intoxicated mice was decreased significantly than that of the controls (66.02 ± 2.31 vs. 104.62 ± 3.62 U/mg protein). Pre-treatment with Poly-herbal medicine (Trasina[®]) significantly increased Serum SOD activity compared with ethanol intoxicated mice (99.65 ± 2.01 vs. 66.02 ± 2.31 U/mg protein).

Hepatic super oxide dismutase (SOD) activity in the ethanol intoxicated mice was significantly less than that of controls (43.85 ± 1.84 vs 86.27 ± 1.59 U/mg protein). Pre-treatment with Poly-herbal medicine (Trasina®) significantly increased (Figure 2) hepatic SOD activity compared with ethanol intoxicated mice (80.02 ± 2.21 vs. 43.85 ± 1.84 U/mg protein).

Protective effect of Poly-herbal medicine (Trasina®) on serum and liver CAT activity in ethanol toxicity

Serum, and liver catalase (CAT) activities are depicted in Figure 3. Serum CAT activity in the ethanol intoxicated mice was significantly less than that of the normal untreated animals (149.26 \pm 6.85 vs. 212.36 \pm 5.98 U/mg protein). Pre-treatment with Poly-herbal medicine (Trasina®) significantly increased Serum CAT activity compared with ethanol intoxicated mice (200.55 \pm 5.71 vs. 149.26 \pm 6.85 U/mg protein).

Hepatic catalase (CAT) activity in the ethanol intoxicated mice was significantly less than that of controls (91.26 \pm 2.64 vs 165.48 \pm 6.22 U/mg protein). Pre-treatment with Poly-herbal medicine (Trasina[®]) significantly increased (Figure 3) hepatic CAT activity compared with ethanol intoxicated mice (162.65 \pm 4.78 vs. 91.26 \pm 2.64 U/mg protein).

Protective effect of Poly-herbal medicine (Trasina®) on serum and liver GSH activity in ethanol toxicity

Serum, and liver reduced glutathione (GSH) activities are depicted in Figure 4. Serum GSH activity in the ethanol intoxicated mice was significantly less than that of the controls (20.20 ± 0.61 vs. 39.67 ± 0.88 U/mg protein). Pre-treatment with Poly-herbal medicine (Trasina[®]) significantly increased Serum GSH activity compared with ethanol intoxicated mice (40.19 ± 1.51 vs. 20.20 ± 0.61 U/mg protein).

Hepatic reduced glutathione (GSH) activity in the ethanol intoxicated mice was significantly less than that of controls (9.06 ± 0.35 vs 18.62 ± 0.84 U/mg protein). Pre-treatment with Poly-herbal medicine (Trasina[®]) significantly increased (Figure 4) hepatic GSH activity

compared with ethanol intoxicated mice (21.05 ± 1.44 vs. 9.06 ± 0.35 U/mg protein).

Protective effect of Poly-herbal medicine (Trasina®) on serum and hepatic Glutathione –S Transferees (GST) activity in ethanol toxicity

Serum and hepatic Glutathione –S Transferees (GST) activities are depicted in Figure 5. Serum GSH activity in the ethanol intoxicated mice was significantly less than that of the controls $(3.11 \pm 1.02 \text{ vs. } 8.32 \pm 0.99 \text{ U/mg} \text{ protein})$. Pre-treatment with Poly-herbal medicine (Trasina®) significantly increased Serum GSH activity compared with ethanol intoxicated mice (8.77 $\pm 1.51 \text{ vs. } 3.11 \pm 1.02 \text{ U/mg} \text{ protein})$.

Tissue level (Liver) Glutathione –S Transferees (GST) concentration in the ethanol intoxicated mice was significantly less than that of controls $(3.18 \pm 0.41 \text{ vs} 6.74 \pm 0.79 \text{ U/mg}$ protein). Pre-treatment with Polyherbal medicine (Trasina®) significantly increased (Figure 5) hepatic GSH activity compared with ethanol intoxicated mice (6.17 ± 0.25 vs. 3.18 ± 0.41 U/mg protein).

DISCUSSION

Chronic intake of alcohol generates reactive oxygen species (ROS) which produced cellular damage in every mammalian species. During ethanol intoxication, liver is the main target organ for oxidative stress. According to the various scientific research different free radicals as Superoxide anion $(0_2^{\bullet-})$, hydroxyl radical (OH $^{\bullet}$), and hydrogen peroxide (H₂O₂) are the major ROS generated during normal redox reaction in our body developed cytotoxic effects. These ROS molecule are generally neutralized by the defensive action of the endogenous antioxidant system, primarily composed of glutathione glutathione superoxide dismutase [28]. [29], peroxidase and catalase [30]. When body lost the equilibrium between ROS production and antioxidant defensive system can create severe oxidative stressinduced damage, consequently, ROS accumulation may cause protein oxidation leading to the disruption of cell membranes, organelles, and loss of function [31].

During the cellular oxidative stress lipid peroxidation is commonly used as marker. During cellular lipid oxidation malondialdehyde (MDA) is generated as an end product denoted as a marker of lipid peroxidation [32]. Treatment of mice with ethanol (50% v/v) significantly elevated lipid peroxidation which is reflected as elevated MDA levels in serum and liver tissues. The condition indicates that cell was severely affected by reactive oxygen species (ROS) leads to cellular damage. Application of poly-herbal formulation (Trasina®) significantly reduced the serum MDA level. The treatment also reduced the hepatic MDA levels which clearly indicate that normal membranous fluid level of the cell maintained by herbal medicine which plays a vital role in cell functioning. Apart from this animals intoxicated with ethanol decreased serum and liver total protein indicated the cellular damage. Simultaneous treatment with Trasina[®] normalized the serum and liver protein level towards experimental animals.

The normal activity of first order anti-oxygen enzymes mainly Super oxide dismutase (SOD), Catalase (CAT) and second order enzymes Glutathione –S Transferase (GST) and Glutathione (GSH) are commonly inhibited by intoxication of ethanol resulting oxidative damage. Sever oxidative stress suppress the normal cellular functions and gradually damage the cellular activity.

To maintain the cellular homeostasis balance between antioxidant defensive system and ROS production is very crucial. In this study our aim is to determine the possible therapeutic effect of poly herbal medicine (Trasina®) upon antioxidant enzymes activities as a marker of oxidative stress. Scientific study reviled that first order antioxidant enzyme i.e. SOD is works against the superoxide radical and catalyzes its dismutation into H₂O₂, which is utilized by catalase (CAT) or glutathione peroxidase (GPx) [33]. On the other hand GST catalyzes the conjugation of several substrates to the thiol group of glutathione, transforming toxic materials into less toxic forms [34,35]. In the present study, oral administration of ethanol (50% v/v) on mice significantly reduced the serum and liver antioxidant enzyme activities as compared to control untreated animals which supported the previous experiment that chronic consumption of ethanol generate free radicals which reduced the antioxidant enzymes activities. Generation of reactive oxygen species (ROS) within the cell decreased the cellular performance by changing the antioxidant enzymes actions. Treatment with poly herbal medicine (Trasina[®]) at a dose of 200 mg/kg/day on mice those are intoxicated with ethanol, significantly elevated serum and liver the SOD, GPx, GST and CAT enzyme activities. From this result it is clear that this herbal medicine (Trasina®) inhibit the free radical production within the cell which indicate that synergistic action of various plants compounds in a single medicine may potent to prevent cellular oxidative stress and boost the cell for their normal function.

CONCLUSION

Consumption of ethanol generates reactive oxygen species (ROS) gradually developed oxidative stress in mammalian system. Chronic administration of ethanol alter serum and liver markers and decline various essential antioxidant enzymes activities. Serum and tissue MDA level, the marker of membrane damage drastically elevated in mice after intoxication with ethanol. This may probably contribute to the additional progression of ethanol intoxication related problems and developed cellular deformities. Treatment with poly-herbal medicine (Trasina[®]) normalized the serum and tissues antioxidant enzymes activities by suppression of extensive ROS generation during ethanol intoxication. Thus Trasina® capsule composed of various medicinal herbs may be a potent drug which sound for prevention of cellular oxidative stress.

CONFLICT OF INTEREST

We declare that we have no conflict of interest. **ACKNOWLEDGEMENT**

The authors are thankful to Prof. (Dr.) T.K.Pal, Department of Pharmaceutical Technology, Jadavpur University, Kolkata-700032 and Prof. S K Pal, Senior Professor Department of Chemical, Biological & Macromolecular Sciences S N Bose National Centre for Basic Sciences JD Block, Sector III Salt Lake City, Kolkata for their valuable suggestions and Mr. Gautam Dey, M.D. & Mr. Ranajit Dey, Jt. M.D. for facilities and encouragement during this investigation.

REFERENCES

- 1. Sies H et al. Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. Nature Reviews Molecular Cell Biology. 2020; 21(7):363-383.
- 2. Huihui Z et al. Toxic effects of heavy metals Pb and Cd on mulberry (*Morus alba L.*) seedling leaves: Photosynthetic function and reactive oxygen species (ROS) metabolism responses. Ecotoxicology and environmental safety. 2020; 195:110469.
- Zorov DB et al. Mitochondrial reactive oxygen species (ROS) and ROS-induced ROS release. Physiological reviews. 2014; 94(3):909-950.
- 4. Yu Z et al. Reactive oxygen species-related nanoparticle toxicity in the biomedical field. Nanoscale research letters. 2020; 15:1-4.
- 5. Goc Z et al. Effect of taurine on ethanol-induced oxidative stress in mouse liver and kidney. Chin J Physiol. 2019; 62:148-156.
- 6. Jing L et al. Chronic alcohol intake-induced oxidative stress and apoptosis: Role of CYP2E1and calpain-1 in alcoholic cardiomyopathy. Mol Cell Biochem. 2012; 359:283-292.
- 7. Simplicio JA et al. Reactive oxygen species derived from NADPH oxidase play a role on ethanol-induced hypertension and endothelial dysfunction in rat resistance arteries. J Physiol Biochem. 2017; 73:5-16.
- Darbar S et al. Protective effect of Livina, a polyherbal liquid formulation against ethanol induced liver damage in rats. Ant sci life. 2009; 28(3):14-17.
- 9. Khodaei F et al. Effect of sodium benzoate on liver and kidney lipid peroxidation and

antioxidant enzymes in mice. J Rep Pharm Sci. 2019; 8:217-223.

- 10. EL-Shenawy NS et al. Mitigating Effect of Ginger against Oxidative Stress Induced by Atrazine Herbicides in Mice Liver and Kidney. J Biofertil Biopestici. 2011; 2:107.
- 11. Shah ZA et al. Antioxidant/restorative effects of calcined gold preparations used in Indian systems of medicine against global and focal models of ischaemia. Pharmacology & toxicology. 2002; 90(5):254-259.
- 12. Mukherjee PK. Exploring botanicals in Indian system of medicine—regulatory perspectives. Clinical Research and Regulatory Affairs. 2003; 20(3):249-264.
- 13. Darbar S et al. Antioxidant and hepatoprotective activity of Livina, a polyherbal liquid formulation. Asian J Chem. 2009; 21(2):1495-1499.
- Cederbaum AI et al. Role of oxidative stress in alcohol-induced liver injury. Arch Toxicol, 2009; 83(6): 519-548.
- 15. Darbar S et al. Single Dose Acute Oral Toxicity of Livina, a Polyherbal Formulation in Mice Model. European Journal of Pharmaceutical and Medical Research. 2018; 5(2): 492-495.
- 16. Darbar S et al. Preliminary Single Dose Toxicological Investigation of Livina Capsule in Mice Model. International Journal of Research in Pharmacy and Pharmaceutical Sciences. 2020; 5(3): 26-28.
- 17. Darbar S et al. Effect of a polyherbal liquid formulation on aceclofenac induced gastric mucosal damage in albino wistar rats. J Pharm Res. 2008; 7(2):62-65.
- Darbar S et al. Anti-Stress Activity (in-vivo) of Multi Herbal Capsule-Trasina® in Experimental Murine Model. Asian Journal of Pharmaceutical Research and Development. 2020; 8(5):52-58.
- 19. Bhattacharya SK et al. Adaptogenic activity of Withania somnifera: an experimental study using a rat model of chronic stress. Pharmacology Biochemistry and Behavior. 2003; 75(3):547-555.
- 20. Darbar S et al. Assessment of Acute Oral Toxicity Study of Trasina®, an Ayurvedic

Herbal Formulation on Experimental Models. J. Pharm. Med. Res. 2019; 4(1): 84–86.

- 21. Darbar S et al. Preliminary acute oral toxicity study of newly developed herbal formulation, World J Pharm Res. 2018; 7(5): 924-930.
- 22. Kashyap M et al. A rapid and simple method for measurement of total protein in very low density lipoproteins by the Lowry assay. J Lipid Res. 1980; 21:491-505.
- 23. Ohkawa H et al. Assay for lipid peroxidase in animal tissue by thiobarbituric acid reaction. Anal Biochem. 1979; 95: 351-358.
- 24. Beutler E. Red Cell Metabolism, a Manual of Biochemical Methods (third ed) Grune and Startton, 1984; New York p 133.
- 25. Misra HP et al. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. J Biol Chem. 1972; 247: 3170-3175.
- 26. Beutler E et al. Improved method for determination of blood glutathione. From the department of medicine, city of hope medical centre, J Lab Clin Med. 1963; 61: 882-888.
- 27. Jollow DJ et al. Bromobenzene-induced liver necrosis. Protective role of glutathione and evidence for 3,4-bromobenzene oxide as the hepatotoxic metabolite Pharmacology. 1974; 11(3):151-169.
- Hsu JY et al. Aqueous Extract of Pepino (Solanum muriactum Ait) leaves ameliorate lipid accumulation and oxidative stress in alcoholic fatty liver disease. Nutrients. 2018; 10: 931.
- 29. Milat AM et al. Effects of white wine consumption on weight in rats: Do polyphenols matter? Oxid. Med. Cell Longev 2017; 1: 1-7.
- 30. Lieber CS. Relationships between nutrition, alcohol use and liver disease. Alcohol Res Health. 2003; 27: 220–231.
- Feinman L et al. Nutrition and diet in alcoholism. In Modern Nutrition in Health and Disease, 9th ed.; Shils, M.E., Olson, J.A., Shike, M., Ross, A.C., Eds.; Williams & Wilkins: Baltimore, MD, USA, 1998; pp. 1523–1542.
- 32. Zeyuan D et al. Effect of green tea and black tea on the blood glucose, the blood triglycerides

and antioxidation in aged rats. J Agric Food Chem. 1998; 46:3875-3878.

- 33. Zararsiz I et al. Protective effect of melatonin against formaldehyde- induced kidney damage in rats. Toxicol Ind Health. 2007; 23:573-579.
- 34. Winterbourn CC. Concerted antioxidant activity of glutathione and superoxide dismutase. In: Packer L, Fuchs J (ed) Biothiols in health and disease. Marcel Dekker Inc, New York; 1995, pp 117-34.
- 35. Abuja PM et al. Methods for monitoring oxidative stress, lipid peroxidation and oxidation resistance of lipoproteins. Clinica chimica acta. 2001; 306(1-2):1-7.

Table 1: Composition of Poly-herbal Formulation (Trasina®)Each capsule contains:Powder and Extractive derived from:

Sl. No.	Scientific Name	Common Name	Family	Quantity
1.	Ocimum sanctum	Tulsi	Lamiaceae	190 mg
2.	Withania somnifera	Ashwagandha	Solanaceae	60 mg
3.	Picrorhiza kurroa	Kutki	Plantaginaceae	10 mg
4.	Eclipta alba	Bhringraj	Asteraceae	10 mg
5.	Tinospora cordifolia	Guduchi	<u>Menispermaceae</u>	10 mg

Table 2: Protective effect of Poly-Herbal Formulation (Trasina®) on Serum and liver total protein content in ethanol intoxicated mice.

Groups	Total Protein (mg/dL)		
	Serum	Liver	
Control	7.52 ±0.24	5.92±0.11	
Ethanol (50% v/v)	3.62±0.16#	2.05±0.09#	
Ethanol + Livina® (200mg/kg)	7.02±0.18*	5.09±0.18*	
Livina® (200mg/kg)	7.41±0.25*	5.51±0.17*	

Values are mean \pm SD of six observations. (n=10) #significant difference from control mice (P \leq 0.001). *significant difference from ethanol intoxicated group (P \leq 0.05).

Table 3: Protective effect of Poly-Herbal Formulation (Trasina®) on Serum and liver MDA level in ethanol intoxicated mice.

Groups	MDA (nmol/g)		
	Serum	Liver	
Control	45.92 ±1.54	35.28±0.92	
Ethanol (50% v/v)	102.58±1.87#	77.05±0.79#	
Ethanol + Livina® (200mg/kg)	49.63±0.91*	39.01±0.88*	
Livina® (200mg/kg)	42.11±0.95*	34.51±0.97*	

Values are mean \pm SD of six observations. (n=10) #significant difference from control mice (P \leq 0.001). *significant difference from ethanol intoxicated group (P \leq 0.05).

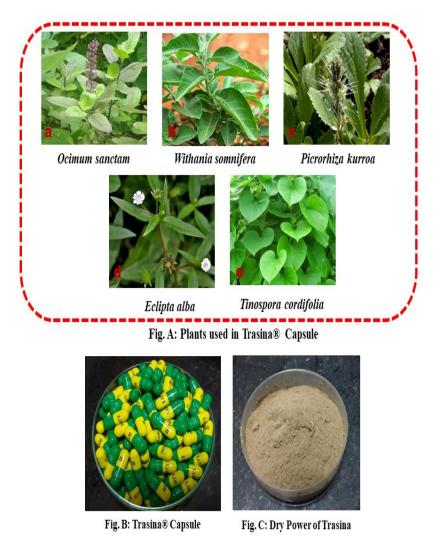


Figure 1: Poly-Herbal medicine – Trasina[®] with different ingredients those are present in the formulation.

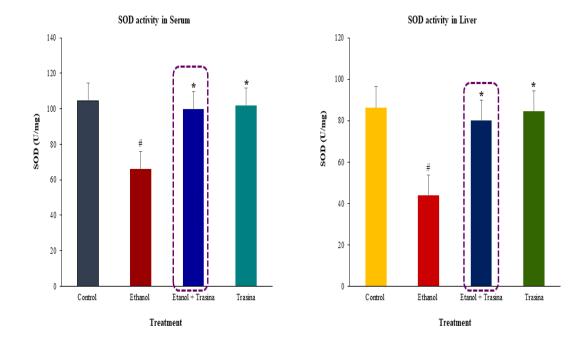


Figure 2: Effect of ethanol alone or in combination with poly-herbal medicine (Trasina[®]) on serum and liver super oxide dismutase (SOD) activity. Values expressed are mean \pm SE (n=10). #significantly different from control group *P* < 0.001 and *significantly different from ethanol treated group *P* < 0.001.

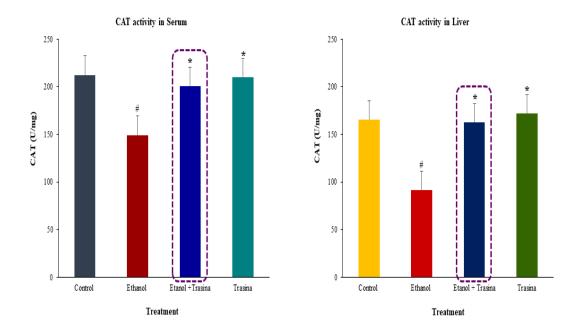


Figure 3: Effect of ethanol alone or in combination with poly-herbal medicine (Trasina[®]) on serum and liver Catalase (CAT) activity. Values expressed are mean \pm SE (n=10). #significantly different from control group *P* < 0.001 and *significantly different from ethanol treated group *P* < 0.001.

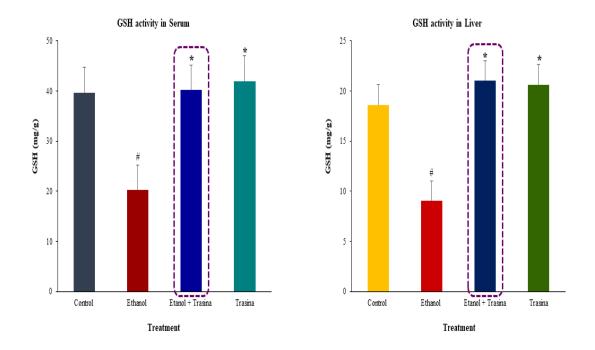


Figure 4: Effect of ethanol alone or in combination with poly-herbal medicine (Trasina[®]) on serum and hepatic glutathione (GSH) activity. Values expressed are mean \pm SE (n=10). #significantly different from control group *P* < 0.001 and *significantly different from ethanol treated group *P* < 0.001.

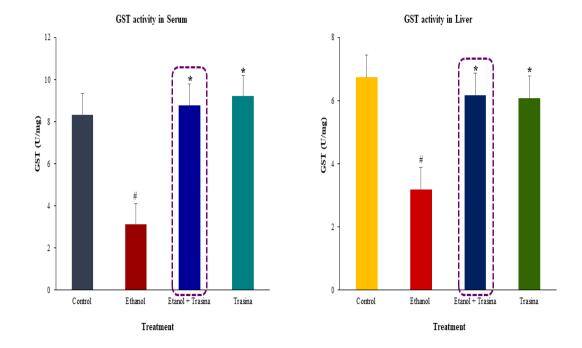


Figure 5: Effect of ethanol alone or in combination with poly-herbal medicine (Trasina[®]) on serum and hepatic glutathione –S transferase (GST) activity. Values expressed are mean \pm SE (n=10). #significantly different from control group *P* < 0.001 and *significantly different from ethanol treated group *P* < 0.001.